## ANTIMICROBIAL AND ANTIOXIDANT ACTIVITIES OF UNRIPE PAPAYA

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### Summary

The meat, seed and pulp of Carica papaya Linn., a popular traditional medicinal herb grown in the tropics, was shown by the agar-cup method to be bacteriostatic against several enteropathogens such as Bacillus subtilis, Enterobacter cloacae, Escherichia coli, Salmonella typhi, Staphylococcus aureus, Proteus vulgaris, Pseudomonas aeruginosa, and Klebsiella pneumoniae. The same parts of papaya were unequivocably demonstrated by electron spin resonance spectrometry to scavenge 1.1diphenyl-2-picrylhydrazyl (5.8 x 10<sup>14</sup> spins/ml), hydroxyl (5.1 x 10<sup>14</sup> spins/ml) and superoxide (1.2 x 10<sup>14</sup> spins/ml) radicals with the seed giving the highest activity at concentrations (IC<sub>50</sub>) of 2.1, 10.0 and 8.7 mg/ml, respectively. The superoxide dismutase (SOD)-like activity in the meat, seed and pulp amounts to about 32, 98 and 33 units/ml; comparable to those of soybean paste miso, rice bran and baker's yeast. Vitamin C, malic acid, citric acid and glucose are some of the possible antioxidative components in papaya. Our study correlates the bacteriostatic activity of papaya with its scavenging action on superoxide and hydroxyl radicals which could be part of the cellular metabolism of such enteropathogens. This is indicative of the pathophysiological role of these reactive oxygen species in gastrointestinal diseases and papaya's ability to counteract the oxidative stress.

The use of plants with bioactive compounds in humans has become one of the most important yet conservative approaches in preventive medicine. Although more than 6,000 antibiotics have been discovered so far from various plant materials (1), none guaranteed absence of side-effect and antibiotic resistance to the host organism-inciting a continuing search for clinically useful natural antimicrobial agents. Few

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studies have linked these antimicrobials as potential antioxidants and scavengers of reactive oxygen species (ROS). The metabolism of enteropathogens caused the production of superoxide and hydrogen peroxide which react, and aided by the presence of iron in feces, efficiently generate hydroxyl radicals. This intracolonic free radical formation may explain the high incidence of cancer in the colon and rectum (2). The same group of researchers have demonstrated that superoxide and hydrogen peroxide generated by activated leukocytes form hydroxyl radicals through Fenton reaction resulting in direct membrane disruption by lipid peroxidation (3). Van der Vliet et al. have also shown that the production of ROS may cause an increased permeability of the endothelium and mucosa, allowing infiltration of inflammatory leukocytes into the ischemic area of the intestine. They proposed that scavenging of ROS will be beneficial in intestinal diseases (4).

Philippine ethnomedical information on *Carica papaya* Linn., a popular traditional medicinal herb, reveals that the fruits, stems, leaves and roots may be used as anthelmintics, stomachic, antidyseptic, diuretics, emmenagogue, laxative, vermifuge, antiasthmatic, antirheumatic, rubefacient, tonic, poultice, and as a cure for enlargement of liver, spleen, freckles and cancerous growths, among others (5).

To further explain scientifically these anecdotal therapeutic cures of papaya, we studied its antimicrobial activity on some gastrointestinal disease-causing enteropathogens and its antioxidant activity. Using unripe papaya, we have previously demonstrated that it exerts antioxidant protection on superoxide, hydroxyl and 1,1-diphenyl-2-picrylhydrazyl radicals (6) in line with other findings (7). In another light, this study was also prompted by the unpublished clinical reports on Bio-normalizer (BN), a fermented food supplement of which a major component is papaya, and its purported therapeutic action on some of the above-mentioned diseases and disorders. BN is a hydroxyl radical scavenger (8) and an inhibitor of neuronal lipid peroxidation in a number of experimental animal models (8-10). We also compared the antioxidant action of the different parts of papaya to its known antioxidative components such as vitamin C, malic and citric acid and glucose. Further, we compared papaya's antioxidant activity to soybean paste miso, rice bran and baker's yeast.

### Materials and Methods

**Plant sample**: A 2-3 week old Philippine variety of *Carica papaya* Linn. (pawpaw) which originated from tropical America, was used. The fresh meat, seed and pulp of papaya and the mixture of these three parts were osterized in a waring blender. The supernatant was used.

Chemicals: DPPH, hypoxanthine, and diethylene-triaminepentaacetic acid (DETAPAC) and SOD (bovine serum; 2,800 units mg<sup>-1</sup>) were obtained from Sigma Chemical Co., St Louis, MO. (USA); xanthine oxidase was from Boehringer, Mannheim (Germany) and 5,5'-dimethyl-1-pyrroline-N-oxide (DMPO) was from Daiichi Pure Chemicals Co., Ltd., Tokyo, (Japan). All other chemicals, reagents and standard culture media used were of the highest grade available from commercial suppliers.

**Microorganisms:** Bacillus subtilis, Enterobacter aerogenes, Enterobacter cloacae, Escherichia coli, Klebsiella pneumoniae, Proteus vulgaris, Pseudomonas aeruginosa, Salmonella typhi, and Staphylococcus aureus, preserved in the Microbiology Section of the University of Santo Tomas Research Center for the Natural Sciences, were originally obtained from the American Type Culture Collection.

**Medium and culture conditions**: Assay medium contained 3.0 g beef extract, 5.0 g peptone, and 2% agar in one liter of distilled water, pH 7.0. A 24-h culture of the microorganisms incubated at 37°C was used in the study.

Screening for antimicrobial property: The inoculum density of the bacterial suspension was standardized at  $3 \times 10^8$  organisms/ml by comparing it with the barium sulfate standard by the turbidimetric method (11). The seeded nutrient agar contained 1% inoculum and all trials were done in triplicates. Agar-cup method was employed for the antibiotic assay.

Analysis of free radicals: The details of the analysis of DPPH, hydroxyl, and superoxide radicals by ESR spectrometry (JEOL, JES-FE1XG, Tokyo, Japan) were given in previous reports (6,8,12-14).

Analysis of SOD: Analysis of SOD activity was performed by ESR spectrometry (9,15). The osteurized papaya juice was centrifuged at 2,000 rpm for 15 min and the supernatant was used. Fifty microliters of 2 mM hypoxanthine, 35  $\mu l$  of 11 mM DETAPAC, 50  $\mu l$  of the supernatant, 15  $\mu l$  of DMPO, and 50  $\mu l$  of freshly prepared xanthine oxidase (81.6  $\mu l$  in 5 ml of phosphate buffer, pH 7.8) were mixed and sucked in a 169  $\mu l$ -volume capacity special flat cell (JEOL, Tokyo, Japan). The spin adduct of superoxide radical (DMPO-OOH) was recorded exactly after 1 min of mixing the xanthine oxidase. A standard SOD curve was prepared from different diluted concentrations of SOD in 0.1 M sodium phosphate buffer (pH 7.4). Manganese oxide was used as an internal standard. The spin number was calculated from the ratio signal height intensity of 2,2,6,6-tetramethyl-4-hydroxyl piperidine-1-oxyl with known spin number. The SOD-like activity was expressed as unit/ml.

Statistical analysis: Statistical analysis was performed using Student's t-test.

## Results

The mixture of unripe papaya comprising the meat, seed and pulp was bacteriostatic at a concentration of 500 mg/ml against several standard cultures of enteropathogens such as *Staphylococcus*, *Escherichia*, *Proteus*, *Salmonella*, *Klebsiella* and *Enterobacter*. Instead of zones of inhibition, thinning of growth was observed and measured at 500 mg/ml. This minimum inhibitory concentration (MIC) for papaya extract also impaired the growth of *Bacillus subtilis* (Table 1).

The same parts of unripe papaya were unequivocably demonstrated by ESR spectrometry to quench DPPH, hydroxyl and superoxide radicals. The IC<sub>50</sub> of antioxidants and food substances with antioxidant properties is summarized in Table 2. Among the pure antioxidants, vitamin C appears to be the most potent on the three radicals followed by vitamin E. Citric acid shows the highest hydroxyl radical scavenging ability while baker's yeast seems to be the best among the natural food antioxidants although papaya was the only one evaluated based on wet weight basis. Table 3 shows the SOD-like activity of the different papaya parts in comparison to other naturally occurring food antioxidants such as soybean paste miso, rice bran and baker's yeast. The seed of papaya gave the highest enzyme activity (based on ESR spectrometry and spin trapping technique) among the different papaya parts and other food antioxidants.

TABLE 1

Antimicrobial activity of a mixture of unripe papaya (Philippine variety).

Organisms	Zone of thinning (in mm)*			
Staphylococcus aureus	10.5			
Escherichia coli	13.5			
Pseudomonas aeruginosa	9.0			
Proteus vulgaris	5.0			
Salmonella typhi	8.0			
Klebsiella pneumoniae	10.5			
Enterobacter cloacae	13.0			
Bacillus subtilis	15.0			

<sup>\*</sup>Values are averages of three trials. The minimum inhibitory concentration (MIC) of papaya used is 500 mg/ml.

TABLE 2

Concentration of antioxidants/scavengers required to eliminate 50% (IC<sub>50</sub>) of the generated free radicals.

Antioxidant/ scavenger (μg/ml)	DPPH (a) spins/ml x 10 <sup>14</sup>			Superoxi spins x10 <sup>14</sup>		Reference
(µg/m)	X 1017	X 10.4	X 1010	X1014	X1015	
Vitamin C	0.80		35		24	1
Vitamin E	2.84		37		*	1
Citric Acid	NR	7.37		NR		This work
Malic Acid	NR	12.64		NR		11
Glucose	NR	50.15		NR		n
Papaya						н
meat	25,000	67,140		114,530		
seed	2,130	10,000		8,730		
pulp	6,830	47,000		16,820		
Soybean paste	5,000	1 51	4,000		50,000	12
Rice bran	875	2,790		3,730		13
Baker's yeast	200		2,040		220	14

Except for papaya, all samples were analyzed in dry weight basis. Data are means of 5-6 determinations. \* Delayed quenching action observed after 6~9 min, in lieu of the usual analytical time of 50 s for hydroxyl radical generation from Fenton reaction. NR = no reaction. (a) DPPH in ethanolic solution; (b) hydroxyl radical generated from Fenton reaction; (c) superoxide radical generated from hypoxanthine-xanthine oxidase system.

TABLE 3

Superoxide dismutase (SOD)-like activity of some naturally occurring antioxidants in food.

Antioxidants	SOD-like activity (unit/ml)*		
Papaya meat	32		
" seed	98		
" pulp	33		
Soybean paste miso	58		
Rice bran	65		
Baker's yeast	75		

Except for papaya, all samples were analyzed in dry state. \*Based on the ability of the food antioxidants at a concentration of 25 mg/ml to dismutate the superoxide radicals generated from hypoxanthine-xanthine oxidase system.

### Discussion

Our data shows that the mixture of meat, seed and pulp of unripe papaya has bacteriostatic property, although weak at 500 mg/ml (based on fresh weight basis of the plant material). against several major human enteric pathogens and Bacillus subtilis (Table 1) through maybe inhibition of either the bacterial cell wall synthesis or protein synthesis. The extract of unripe papaya is most sensitive to *Escherichia*, *Enterobacter and Klebsiella*. In addition to impairing the growth of these In addition to impairing the growth of these enteropathogens, the different parts of unripe papaya were found to scavenge various radicals such as DPPH, hydroxyl and superoxide. As part of the bacterial metabolism. ROS such as superoxide and hydrogen peroxide are generated and in the presence of hemoglobin-, blood- and feces- containing iron may produce hydroxyl radicals. The ensuing oxidation disrupts the membrane through lipid peroxidation. Colonization, penetration and proliferation of bacteria within the intestinal and mucosal epithelium may cause lesions, abscesses, bleeding, secretion of toxins and production of ROS. Stopping or halting the growth of these enteropathogens through papaya's bacteriostatic action corollarily implies inhibition of the production of ROS thereby arresting or slowing down the progression of such gastrointestinal diseases. There is convincing evidence to suggest that ROS are generated in various gastrointestinal pathologies such as intestinal ischemia and reperfusion (16), hemorrhagic shock (17), gastrointestinal inflammatory diseases (18), acute pancreatitis (19), inflammatory bowel diseases (20), xenobiotic metabolism (21), colon cancer (2), and ulcerative colitis (3), among others. The scavenging of ROS at this stage appears beneficial in these as the status of the oxidation-reduction potential of the intestinal microflora is maintained. In the case of unripe papaya, its antioxidant property (IC50)is more than ten-fold higher than its antimicrobial activity (MIC). While all antimicrobials have limitations on their antimicrobial spectra, the antioxidants are far-reaching and may be beneficial in the prevention of free radical-induced diseases including those of microbial origin.

Daley et al. has characterized the low molecular weight protein components of papaya, which are similar to purothionins, as heat- and acid-tolerant and have physiological functions related to redox activity (22). Benzyl isothiocyanate, a mustard oil produced by enzymatic hydrolysis of benzyl glucosinolate by thioglucosidase in

papaya (23) is suggested to possess antimicrobial activity. The latex of immature fruit of papaya which is used as amebicide in Central America (24) has the proteolytic Papaya's antioxidative enzyme papain and potassium benzyl glucosinolate (23). components are attributable to its vitamin C content, malic acid, citric acid and glucose (Table 2). The SOD-like activity of unripe papaya, which provides the first line of antioxidant defenses against superoxide radicals, is comparable to those of baker's yeast, soybean paste miso and rice bran (Table 3). While its SOD-like activity has yet to be purified, it is worth noting that SOD compounds have been purified from a number of plant origins, namely: Fe-SOD from tomato and mustard; Mn-SOD from maize and pea; and Cu,Zn-SOD from green pea, maize, wheat germ, spinach chloroplast, pea and corn seedlings, cucumber and green pepper (25). The monosaccharides-, saccharose. levulose- and levulinic acid- contents of papaya may also contribute to its antioxidant property (5,6,14,25). These antioxidants in papaya may in part explain why BN which has papaya as its base material is a potent hydroxyl radical scavenger and inhibitor of lipid peroxidation (8-10). In addition, BN was found effective in stopping the growth of several facultative isolates from 84 gastrointestinal specimens of patients suffering from gastric ulcer, amoebic colitis, diarrhea and hemorrhoids (unpublished data).

Our study correlates two significant pharmacological activities of unripe papaya, as a bacteriostatic agent and as an antioxidant. The study further suggests that papaya may be an important inexpensive clinical agent on gastrointestinal problems of microbial

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# References Programme Progr

- S. OMURA. Microbiol. Revs. 50 259-279 (1986).
- C.F. BABBS. Free Radical Biol. Med. 8 191-200 (1990).
- C.F. BABBS. Free Radical Biol. Med. <u>13</u> 169-181 (1992). 3.
- A. VAN DER VLIET. Free Radical Biol. Med. 12 499-513 (1992). 4.
- E. QUISUMBING, Medicinal Plants of the Philippines, 632-635, 5. Katha Publishing, Manila, Philippines (1978).
- L.A. SANTIAGO, J.A. OSATO and A. MORI. Neurosciences 18 189-194 (1992).
- 7. E.J. WEBMAN, G. EDLIN and H.F. MOWER. Intl. J. Radiat. Biol. 55 347-351 (1989).
- 8. L.A. SANTIAGO, J.A. OSATO, M. HIRAMATSU, R. EDAMATSU and A. MORI. Free Radical Biol. Med. 11 379-383 (1991).
- L.A. SANTIAGO, J.A. OSATO, H. KABUTO and A. MORI. Med.Sci. Res. 21 139-9. 141 (1993).
- L.A. SANTIAGO, J.A. OSATO, J. LIU and A. MORI. Neurochem.Res. 18 10. 711-717 (1993).
- O.C. GROVE and W.A. RANDALL, Assay Methods of Antibiotics: A Laboratory 11. Manual, p. 238, Medical Encyclopedia, Inc., New York (1955).
- L.A. SANTIAGO, M. HIRAMATSU and A. MORI. J. Nutr. Sci. Vitaminol. 38 297-12. 304 (1992).

- 13. L.A. SANTIAGO and A. MORI. Med.Sci. Res. 20 827-828 (1992).
- 14. L. A. SANTIAGO, M. HIRAMATSU and A. MORI. Med.Sci. Res. 19 867-868 (1991).
- 15. M. HIRAMATSU, M. KOHNO, K. MITSUTA and A. MORI, J. Neurochem. 58 1160-1164 (1992).
- J.M. McCORD. N. Engl. J. Med. 312 159-163 (1985). 16.
- 17. M. ITOH and P.H. GUTH. Gastroenterology 88 950-956 (1985).
- 18. J.M. BRAGANZA and H. RINDERKNECHT. Gastroenterology 94 1111-1112 (1988).
- M.H. SCHOENBERG, M. BUCHLER, M. GASPAR, A. STINNER, M. YOUNES, I. 19. MELZNER, B. BULTMANN and H.G. BEGER. Gut <u>31</u> 1138-1143 (1990). S.J. WEISS. Acta Physiol. Scand. Suppl. <u>548</u> 9-37 (1986).
- 20.
- 21. C.M. MANSBACH II, G.M. ROSEN, C.A. RAHN and K.E. STRAUSS. Biochim. Biophys. Acta 888 1-9 (1986).
- 22. L.S. DALEY and L.J. THERIOT. J. Agric. Food Chem. 35 680-687 (1987).
- 23. C.S. TANG. Phytochemistry 12 769-773 (1973).
- 24. W.H. LEWIS and M.P.F. LEWIS, Medical Botany, p 292, John Wiley & Sons, New York (1977).
- 25. B. HALLIWELL and G.M.C. GUTTERIDGE (Eds). Free Radicals in Biology & Medicine, Clarendon Press, Oxford (1989).

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